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Introduction: Foot and mouth disease (FMD) is the most contagious disease of cloven-hoofed animals, and has a great potential for causing severe economic loss. Detection of antibodies to non-structural protein (NSP) has been used to identify past or ongoing infections with any of the existing serotypes of FMDV. Surveillance activity consists of passive epidemiological surveillance system of investigating reported disease activity and active epidemiological surveillance system of examining statistically selected and targeted samples within host populations. In this study, we have summarized serological surveillance results for the detection of NSP antibody across the country in 2014.

Materials and Methods: A total of 37,336 sera from 6,965 farms were tested for statistical surveillance. A total of 93,893 sera from 4,613 farms were tested for targeted surveillance. Those sera were tested by NSP ELISA (Median Diagnostics or Bionote) according to the manufacturer's instructions.

Results: A total of 33 samples from 20 farms were NSP antibody positive with 28 cattle, 1 pig and 4 goats in statistical serological surveillance. Targeted serological surveillance found out 127 samples NSP antibody from 18 farms. When we traced back those NSP antibody positive farms, any evidence of virus circulation was not found. In addition, all the samples from 617 wild animals such as wild boars and so on were negative by the NSP ELISA.

Conclusions: The animals with NSP antibody positives did not show any clinical sign and all negative by PCR for antigenic samples.

References

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Characterization of Monoclonal Antibodies Against VP2 and VP4 of Foot-and-Mouth Disease Virus

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Introduction: Foot-and-mouth disease (FMD) is one of the most highly contagious diseases of cloven-hoofed animals including cattle, swine, sheep and goats. The causative agent is a FMD virus (FMDV), which is classified into seven serotypes; O, A, Asia1, C, South African Territory1 (SAT1), SAT2 and SAT3. FMDV is surrounded by an icosahedral capsid that comprises 60 copies of

four structural proteins: VP1, VP2, VP3 and VP4. As a serological method, virus neutralization test (VNT) is a standard test. It, however, takes several days to complete and requires a containment facility. These aspects make the VNT inappropriate for mass serological surveillance. The other prescribed test at OIE manual is the liquid-phase blocking (LPB) ELISA which requires training and experience to yield reproducible results and use an inactivated FMDV as a diagnostic antigen. To address these limitations, recombinant proteins derived from FMDV P1 precursor and 3C protease genes of FMDV type O was developed in our previous study. In this study, monoclonal antibody was developed against VP2 and VP4 of FMDV. We suppose that these mAbs can be applied to characterization of FMD recombinant antigens.

Materials and Methods: VP2, VP4 genes were cloned into Rhamnose SUMO vector separately and expressed recombinant proteins using Rhamnose SUMO Cloning Expression System in BL21 DE3 *E. coli* strain. Soluble VP2 and VP4 proteins were purified using Ni-NTA affinity chromatography and the purified proteins were confirmed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). MAb against VP2 and VP4 were produced by immunizing mouse (Balb/c) with each recombinant VP2 and VP4 proteins.

Results: VP2 and VP4 proteins were purified using Ni-NTA affinity chromatography and the purified proteins were confirmed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). To obtain mAbs, the purified proteins VP2 and VP4 were immunized to mouse followed by cell fusion, and selection according to the general mAb production protocol. As a result, we screened 6 of VP2 and 1 of VP4 monoclonal antibodies using ELISA and fluorescence analysis (FA) and also characterized using isotyping kit respectively.

Conclusions: The recombinant VP2 and VP4 proteins were expressed using Rhamnose SUMO Cloning Expression System and monoclonal antibodies against VP2 and VP4 were also produced with the recombinant proteins. We suppose that these mAbs can be applied to characterization of FMD recombinant antigens.

References

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